

RESEARCH

Open Access



# Acute, sub-acute and developmental toxicity studies of molybdenum disulfide nanoflowers in rats, as per OECD guidelines

Farina Hanif<sup>1</sup>, Aslam Khan<sup>1,2\*</sup>, Fareeha Anwar<sup>1</sup>, Haseeb ullah<sup>3</sup>, Muhammad Furqan Akhtar<sup>1</sup> and Muhammad Imran Khan<sup>1</sup>

## Abstract

**Background** This study aimed to investigate the potential toxic effects of Molybdenum disulfide nano-flowers (MoS<sub>2</sub> NF), which have been suggested as a chemotherapeutic agent, but lack previous toxicity studies.

**Methods** Acute, sub-acute and developmental toxicity studies were conducted following OECD guidelines 425, 407 and 414, respectively.

**Results** In the acute toxicity study, female Wistar rats received logarithmic doses (1.75–550 mg/kg) of MoS<sub>2</sub> NF over 14 days. Results indicated a decrease in oxidative stress markers (CAT, SOD and GSH) and increased MDA levels, along with significant decrease in organ weight compared to normal control. Alterations in liver enzymes, CBC profile and lipid profile and histopathological analysis were observed in MoS<sub>2</sub> NF groups. Sub-acute toxicity (28-day at 3 and 10 mg/kg in both male and female rats) resulted in increased levels of ALT and AST, decreased levels of CAT, SOD and GSH and increased MDA and urea levels. Sperm analysis in male group showed increased motility and concentration, with more defective morphology. In developmental toxicity studies, a 10 mg/kg dose for 21 days decreased all oxidative markers except MDA, which increased. Fetal crown-to-rump length increased, while uterine SOD, CAT and GSH levels decreased. Histopathology revealed organ damage in both sub-acute and developmental studies. Maternal weight remained unaffected, whereas fetal weight showed an increased.

**Conclusion** MoS<sub>2</sub> NF exhibited mild-to-moderate toxicity, however, long-term studies are recommended to assess the safety and therapeutic potential of MoS<sub>2</sub> NF.

**Keywords** Acute toxicity, Sub-acute toxicity, Developmental toxicity, In vivo, Nano-flowers

\*Correspondence:

Aslam Khan  
aslam.khan@riphah.edu.pk; aslamkhan\_mkd@yahoo.co.uk

<sup>1</sup>Riphah Institute of Pharmaceutical Sciences, Riphah International University, Raiwind Campus, Lahore, Pakistan

<sup>2</sup>Riphah Institute of Pharmaceutical Sciences, Riphah International University, Islamabad, Pakistan

<sup>3</sup>Department of Chemistry, Khushal Khan Khattak University, Karak, Pakistan



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>.

## Introduction

The process of transforming matter to an extent near to the atomic level to produce innovative structures, materials, and devices is known as nanotechnology [1]. Nanoparticles can readily penetrate the skin because of their tiny size and accessibility to sensitive organs. The size, morphology, charged surface, and surface modifications can all have an impact on how pathogenic they are. Their adverse effects are mostly caused by the generation of reactive oxygen species, which results in cell death, damage to cells, and reduced function [2].

Molybdenum belongs to second row transitional metals having molecular weight 95.94 and is represented symbolically by Mo [3]. Pure metallic molybdenum does not occur naturally. In solution, molybdenum readily becomes polymerized to form an array of intricate poly-molybdate compounds [4].

Molybdenum disulfide, also known as molybdenite, is the primary source of molybdenum.  $\text{MoS}_2$  levels in mineral deposits are typically quite low, although the flotation process can concentrate them. Nowadays, molybdenum disulfide is mainly generated as a byproduct of copper extraction [5]. Nanoparticles of molybdenum disulfide are utilized for the identification and examination of biomolecules and hazardous chemicals [6]. Recent research has indicated that  $\text{MoS}_2$  may pose a risk under certain biological conditions, particularly when used as imaging agents or drug carriers are bio-distributed to the liver.  $\text{MoS}_2$  causes dose-dependent cytotoxicity in liver kupffer cells [7]. We have synthesized and characterized the  $\text{MoS}_2$  NF and have studied its therapeutic potential in breast cancer [8]. However, its toxicity has not yet been evaluated. Therefore, this study aimed to provide a comprehensive understanding of the toxicological profile of  $\text{MoS}_2$  NF through acute, sub-acute and developmental toxicity studies in Wistar rats and inform safe usage guidelines for their potential applications in medicine and industry.

## Material and methods

### Ethical committee

All the procedures and techniques used in this study were approved by the Ethical committee (REC/RIPS/LHR/2023/068) of Riphah International University, Raiwind Campus, Lahore, Pakistan.

### Study design

Albino wistar rats of weight range 100–160 g (6–7 weeks old) were chosen for conducting the toxicity studies according to the OECD guidelines. The animals were purchased from the University of Veterinary and Animal Science's animal house. For acute toxicity study conducted on female rats, main test was the choice using a onetime dose and observing for 14 days. Sub-acute toxicity study

was conducted by dividing the animals into three groups separate from both sex rats ( $n=5$  for each group) group-I; the control group (CMC), group-II the treatment group at dose 3 mg/kg  $\text{MoS}_2$  NF and group-III being dosed at 10 mg/kg  $\text{MoS}_2$  NF. As there are no in-vivo study standards available for  $\text{MoS}_2$  NF so we use Cisplatin (the only anti-cancer registered metal compound) as standard to choose the dose for the current study. This study was conducted on both the male and female rats for 28 days orally with a once daily dose. Developmental toxicity study was designed by dividing the animals into 2 groups ( $n=5$  for each group); the control group and the treatment group (10 mg/kg  $\text{MoS}_2$  NF dose). Animals in this study group were treated for 19–21 days based on their pregnancy cycle.

### Drugs and chemicals

Molybdenum disulfide, carboxymethylcellulose (CMC), monobasic potassium phosphate, dibasic potassium phosphate, Pyrogallol, hydrogen peroxide ( $\text{H}_2\text{O}_2$ ), trichloroacetic acid, sodium hydrogen carbonate, monobasic sodium phosphate, DTNB (5,5-dithio-bis-2-nitrobenzoic acid), hydrochloric acid, dibasic sodium phosphate, thiobarbituric acid (TBA), Sodium hydroxide, sodium potassium tartarate, copper sulfate, folin phenol. All the chemicals were of analytical grade and purchased from local vendors dealing with the companies.

### Acute toxicity study

OECD guidelines 425 were followed to conduct this study. As the toxicity profile of the drug was not available, so directly main test was conducted. Female rats were given a dose of 1.75 mg as per kg of the body weight. The rats were keenly noticed for first 30 minutes and kept under observation for next 14 days. All the animals survived so new animals were given the next dose of 5.5 mg/kg and the procedure continued till 550 mg/kg (following the log doses of 17.5, 55, 175 and 550 mg/kg) as no animal showed sign of mortality. The animals were sacrificed after 14-days to analyze different parameters [9].

### Sub-acute toxicity study

This study was conducted under OECD guidelines 407. Rats of both sex (male and female) were used. Animals were divided into six groups ( $n=5$  for each group). Group I -The female control group was given 0.5% CMC. Group II (female) received molybdenum disulfide ( $\text{MoS}_2$ ) NF 3 mg/kg and Group III (female) was being treated with 10 mg/kg  $\text{MoS}_2$  NF. Group IV- male control group treated with 0.5% CMC and Group V (male) was given a dose of 3 mg/kg  $\text{MoS}_2$  NF. Group VI (male) treated with 10 mg/kg  $\text{MoS}_2$  NF. The dosing was continued orally for 28 days daily. After completion of the dosing period, blood samples were collected from each group for CBC,

LFTs, RFTs and lipid profiling. Animals were dissected and different organs were removed for analysis of anti-oxidant activity and Histopathology study [10–12].

### Developmental toxicity

Developmental toxicity study was designed according to OECD guidelines 414. Male and female rats were kept for mating until a vaginal plug appeared. The pregnant rats were then divided into two groups (control and treatment group), with  $n=5$  for each group. Treatment group received 10 mg/kg MoS<sub>2</sub> NF daily for 19–21. Following that time period C- section was performed a day before the completion of gestational period carefully collecting the fetus to check different vitals including weight of fetus, no. of live and dead fetus, crown to rump length, total no. of fetus, etc.

### Soft tissue examination

Fetuses were dipped in the saturated solution of picric acid in alcohol (Bouin's solution) for 10–15 days. Following that time period, the fetus got hardened, its skin was removed carefully and all vital organs were removed and viewed individually for any visual abnormality. Each tissue's length and width were measured as well [10].

### Sperm analysis

All the rats were killed by a dislocation of the cervical spine. Castration method was used to remove the testis. Testicles were collected by making an excision on the prescrotal line. Cauda epididymis thus removed was transferred to petri plate having 1 ml of 7.4 pH sodium phosphate buffer. Sperm cells were diffused into the buffer at 37°C. The sperm suspension thus made was used to measure different parameters, including sperm count, its morphology, DNA integrity (using Acridine Orange test) and sperm motility [13, 14].

### Separation of plasma

After the completion of the experimental protocol, blood samples of experimental rats were collected in the EDTA tube by cardiac puncture. Blood samples were centrifuged for 15 minutes at 4000 rpm to separate the blood plasma for biochemical analysis [15].

### Biochemical analysis

The blood plasma that was separated by centrifugation was used for analysis of ALT, AST, ALP levels, Bilirubin, total proteins, globulins, albumins levels in the liver. Renal function tests (RFTs) including creatinine and urea and analysis of lipid profile that included cholesterol, low-density lipoproteins (LDL), high-density lipoproteins (HDL), very-low-density lipoproteins (VLDL) and tri-glycerides using particular kits for each parameter [16].

### Tissue homogenate preparation

After dissecting all the test animals, desired organs (liver, heart, kidney, lungs, brain, ovaries and testis respectively from female and male animals) were removed and weighed. Sodium phosphate buffer was used in ratio 1:10 to prepare homogenates which were centrifuged at 6000 rpm, 4°C temperature for 10 minutes. A clear supernatant was collected and used for the analysis of different parameters [17].

### Oxidative stress markers analysis

Tissue homogenates were used for the analysis of oxidative parameters that are listed below:

#### Assessment of superoxide dismutase (SOD) level

To measure the anti-oxidant SOD levels, 0.1 ml of pyrogallol solution, 0.1 mL of tissue homogenate and 2.8 ml of 0.1 M Potassium phosphate buffer (pH 7.4) were mixed and the optical density of the mixture were recorded at a wavelength of 325 nm. The formula given below was used to calculate the results [18].

$$Y = 0.0095x + 0.1939$$

#### Catalase activity (CAT)

All the three constituents, including 0.05 ml of tissue homogenate, 1.95 mL of 50 mM Phosphate buffer, and 1 mL of H<sub>2</sub>O<sub>2</sub> solution were mixed together to measure the absorbance of reaction mixture at a wavelength of 240 nm. Using the following equation, CAT was calculated [19]

$$CAT = \frac{S.O.D.}{E \times \text{volume of sample} \times \text{mg of protein}}$$

Where; S.O.D = absorbance measured, E = 0.071 [20].

#### Malondialdehyde (MDA) estimation

1 ml of the tissue supernatant, 1 ml of TBA, 1 ml of 15% TCA and 1 ml of HCL were all mixed. The whole reaction mixture was placed for 15 minutes in a hot water bath. It was then chilled in an ice bath and centrifuged later on at 3500 rpm for 10 minutes. Supernatant was taken to measure absorbance at 532 nm wavelength. MDA was then calculated by the formula.

$$\text{Conc. of MDA} = \frac{\text{abs.} \times 100 \times V_t}{1.56 \times 10^5 \times W_t \times V_u}$$

Where; V<sub>u</sub> = Aliquot volume, V<sub>t</sub> = total mixture volume, W<sub>t</sub> = weight of dissected organ [17, 20].

### Estimation of protein levels

To perform this test, reagent A, B, C were prepared to make reagent 1, where Reagent A: 2% sodium carbonate ( $\text{Na}_2\text{CO}_3$ ) in sodium hydroxide (0.1 N), Reagent B: Sodium potassium tartrate 1 % in 100 ml  $\text{H}_2\text{O}$  and Reagent C: 0.5%  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$  in 100 ml water.

Reagent-I: 48 ml of reagent A, 1 ml of B and 1 ml of C were combined to make reagent-I.

Reagent-II was prepared by mixing 1 part folin phenol (2 N) in part of water. 0.2 ml of the tissue homogenate taken and distilled water was added to make it 1 ml. In this diluted homogenate, 4.5 ml of reagent-I was added to incubate for 10 minutes. Reagent-II (0.5 ml) was added to the mixture after incubation and was incubated again for 30 minutes. Absorbance of the sample was observed at a wavelength of 660 nm. Calculate results using the formula:

$$Y = 0.0744x + 0.0087$$

### Estimation of GSH (glutathione) levels

1 ml of supernatant of the tissue homogenate, 1 ml of TCA (10%) was added in 4 ml of phosphate buffer and 0.5 ml of DTNB and mixed well. Precipitates were formed; the supernatant was removed to estimate the absorbance of the sample at 412 nm. Results were then calculated using the equation:

$$\text{GSH level} = \frac{Y - 0.00314}{0.034} \times \frac{DF}{B_t \times V_v}$$

Where; Y=absorbance of sample, DF=dilution factor,  $B_t$ =organ weight calculated,  $V_v$ =volume of homogenate used [20].

### Histopathological analysis

Histopathology of some selected organs, including kidney, lungs, liver, brain, heart, ovaries, testis, were analyzed by fixing with neutral buffered formalin (10%), processed and embedded in the paraffin. The tissues were then sliced into 3–4  $\mu\text{m}$  sections in longitudinal direction. Eosin and hematoxylin dyes were then used to stain

the organ slices. Histology of the slides formed was then studied using YJ-2005DN digital articulated free binocular microscope having 1.30 MP camera [21].

### Statistical analysis

Mean  $\pm$  SEM was used to express all the calculated values. T-test and One-way analysis of variance (ANOVA) were used to examine all the treatment outcomes before a post-hoc Dunnett's test was run to investigate the entire test procedures used.

## Results

### Effect of $\text{MoS}_2$ NF on body and organ weights in acute, sub-acute and developmental toxicity studies

Body weights were found to be parallel to normal control in each study group for acute, sub-acute and developmental toxicity (Supplementary data; Table 1–3). In acute toxicity study, organ weights of brain, lungs, liver and ovary at a high dose of 175 mg/kg showed a significant increase as compared to control, as shown in Table 1. In sub-acute toxicity study, weight of all organs in the male treatment group at both dosage levels of 3 and 10 mg/kg were significantly increased while in females, weight of only liver and lungs were increased as compared to control group (Table 2).

### Effect of $\text{MoS}_2$ NF on CBC profile in acute, sub-acute and developmental toxicity studies

CBC profile of the acute toxicity study after 28 days of dosing showed a significant increase ( $p < 0.001$ ) in Hb at higher doses, while WBCs and leukocytes were increased at low doses. Lymphocytes were significantly ( $p < 0.001$ ) decreased (Table 3). Sub-acute and developmental toxicity studies showed a significant increase ( $p < 0.001$ ) in platelets and lymphocytes count while the neutrophils were decreased ( $p < 0.01$ ) in both studies as compared to normal control (Table 4).

### Effect of $\text{MoS}_2$ NF on oxidative stress markers in acute and sub-acute and developmental toxicity studies

In acute toxicity study, CAT showed decrease levels in all the selected organs except ovary, which showed

**Table 1** Effect of treatment on organ weight of animals calculated as per 100 g of animal weight in acute toxicity studies at different doses of  $\text{MoS}_2$  Nano flowers

Organs	Control group	$\text{MoS}_2$ NF					
		1.75 mg/kg	5.5 mg/kg	17.5 mg/kg	55 mg/kg	175 mg/kg	550 mg/kg
Brain	1.08 $\pm$ 0.062	1.75 $\pm$ 0.068***	1.60 $\pm$ 0.046***	1.70 $\pm$ 0.08***	1.907 $\pm$ 0.049 ***	1.457 $\pm$ 0.083 ***	1.38 $\pm$ 0.09*
Heart	0.437 $\pm$ 0.04	0.63 $\pm$ 0.038**	0.457 $\pm$ 0.03	0.57 $\pm$ 0.029	0.44 $\pm$ 0.029	0.57 $\pm$ 0.02	0.53 $\pm$ 0.029
Liver	3.78 $\pm$ 0.14	6.58 $\pm$ 0.15***	2.63 $\pm$ 0.22***	7.15 $\pm$ 0.518***	7.35 $\pm$ 0.28***	7.51 $\pm$ 0.27***	5.85 $\pm$ 0.19***
Lung	0.82 $\pm$ 0.07	1.43 $\pm$ 0.06**	1.07 $\pm$ 0.049	2.33 $\pm$ 0.10***	2.57 $\pm$ 0.16***	2.45 $\pm$ 0.11***	2.06 $\pm$ 0.07***
Ovary	0.084 $\pm$ 0.009	0.09 $\pm$ 0.01	0.09 $\pm$ 0.012	0.134 $\pm$ 0.02	0.086 $\pm$ 0.009	0.219 $\pm$ 0.021***	0.125 $\pm$ 0.018
Kidney	0.74 $\pm$ 0.05	1.13 $\pm$ 0.03*	0.77 $\pm$ 0.06	1.423 $\pm$ 0.13***	1.07 $\pm$ 0.07*	0.99 $\pm$ 0.06	1.077 $\pm$ 0.072*

Data are showed as mean  $\pm$  SEM, \*\* $p < 0.01$ , \*\*\* $p < 0.001$  as compared with normal control. \*indicates increase in weight

**Table 2** Effect of treatment on organ weights of Male and Female rats calculated per 100 g of animal weight at different doses of MoS<sub>2</sub> NF in sub-acute toxicity studies

Organs	Female			Male		
	Control group		MoS <sub>2</sub> NF	Control group		MoS <sub>2</sub> NF
	3 mg/kg	10 mg/kg		3 mg/kg	10 mg/kg	
Brain	2.567±0.127		2.933±0.156	2.94±0.107	2.770±0.15	3.343±0.143
Heart	1.00±0.064		1.103±0.067	1.047±0.081	0.767±0.07	0.917±0.105
Liver	9.92±0.18		11.81±0.13***	12.61±0.23***	6.82±0.23	11.47±0.26***
Lungs	2.137±0.27		3.99±0.24**	4.82±0.326***	2.117±0.102	3.13±0.13**
Ovary	0.289±0.107		0.250±0.093	0.147±0.029	2.96±0.061	4.44±0.121***
Kidney	1.743±0.142		1.110±0.097*	1.237±0.137	1.087±0.068	2.650±0.144***

Data are expressed as mean±SEM, n=5, \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 as compared with control. \*indicates an increase in animals weight

**Table 3** Effect of MoS<sub>2</sub> Nano flowers on complete blood count (CBC) in acute toxicity studies at different logarithmic doses

Parameters	Unit	Normal control	MoS <sub>2</sub> NF					
			1.75 mg/kg	5.5 mg/kg	17.5 mg/kg	55 mg/kg	175 mg/kg	550 mg/kg
Hemoglobin	g/dL	10.3±0.1	8.60±0.1 <sup>▲▲</sup>	8.30±0.1 <sup>▲▲</sup>	7.80±0.1 <sup>▲▲</sup>	14.0±0.1***	12.0±0.1***	13.5±0.1***
RBCs	×10 <sup>12</sup> /L	5.650±0.33	4.90±0.11	4.61±0.190	4.07±0.03 <sup>▲</sup>	7.31±0.31**	6.75±0.25*	7.85±0.25**
Neutrophils	%	34.650±2.15	26.50±1.70	54.05±1.55**	42.20±1.4	23.0±3.0 <sup>▲</sup>	84.0±1.0***	61.5±4.50***
Lymphocyte	%	66.60±1.60	63.10±2.20	39.0±2.0 <sup>▲▲</sup>	55.25±0.95 <sup>▲</sup>	74.50±1.5	13.0±3.0 <sup>▲▲</sup>	40.0±2.0 <sup>▲▲</sup>
WBCs	×10 <sup>9</sup> /L	3.60±0.10	3.30±0.20	6.65±0.150***	10.40±0.20***	5.30±0.1***	4.60±0.20*	3.35±0.15
Platelet count	×10 <sup>9</sup> /L	632.5±17.5	470.0±10.0	509±11.0	565.0±15.0	626.0±7.50	737.0±8.0	846.5±5.50

Data are expressed as mean±SEM, \*P<0.05, \*\*P<0.01, \*\*\*P<0.001 as compared with control. \*indicates increase levels while ^indicate decreased levels

**Table 4** Effect of MoS<sub>2</sub> NF on CBC profile of rats group in sub-acute and pregnant rats in developmental toxicity studies

Parameters	Units	Sub-acute toxicity				Developmental toxicity		
		Male		Female		Control	MoS <sub>2</sub> NF	Control
		Control	MoS <sub>2</sub> NF	3 mg/kg	10 mg/kg			
Hemoglobin	g/dL	15.37±0.51	14.16±0.92	13.60±0.81	10.96±1.4	12.23±0.77	14.56±1.10	14.56±1.10
RBCs	×10 <sup>12</sup> /L	6.77±0.26	7.3±0.49	7.36±0.42	4.67±0.38	6.26±0.31	7.5±0.5**	5.00±0.89
WBCs	×10 <sup>9</sup> /L	9.07±1.3	8.40±1.09	10.6±1.27	4.20±1.3	6.5±1.26	3.92±0.97	807.3±63.04
Platelet count	×10 <sup>9</sup> /L	402.6±9.1	850.6±42.67 ***	533.33±28.6 *	752.33±7.2	656.3±29.9	836.6±37.8	4.66±0.63
Neutrophils	%	66.66±4.4	7.33±0.88 <sup>▲▲</sup>	10.33±1.45 <sup>▲▲</sup>	29.27±0.65	10.66±1.2 <sup>▲▲</sup>	25.0±1.15 <sup>▲</sup>	48.33±3.38
Lymphocytes	%	39.66±2.6	90.0±2.3***	85.33±2.02***	64.0±2.6	82.3±4.8*	70.0±3.46	40.00±2.64

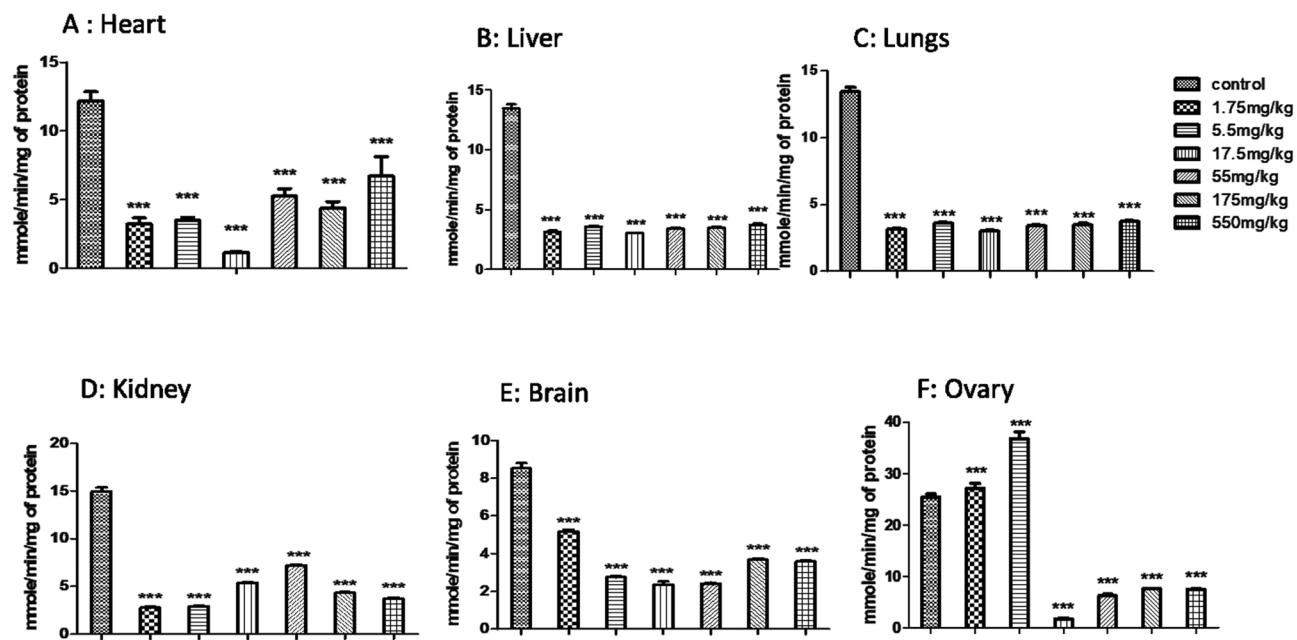
Data are represented as mean±SEM, n=5, \*P<0.05, \*\*P<0.01, \*\*\*P<0.001 as compared with normal control. \*indicates increase levels while ^indicate decreased levels

increased levels at low doses and decrease at high doses (Fig. 1). Glutathione (GSH) (Fig. 2) and superoxide dismutase (SOD) levels were also decreased (Fig. 3) at all doses. MDA levels showed increased activity in all organs except brain that showed significant MDA levels when compared to control (Fig. 4).

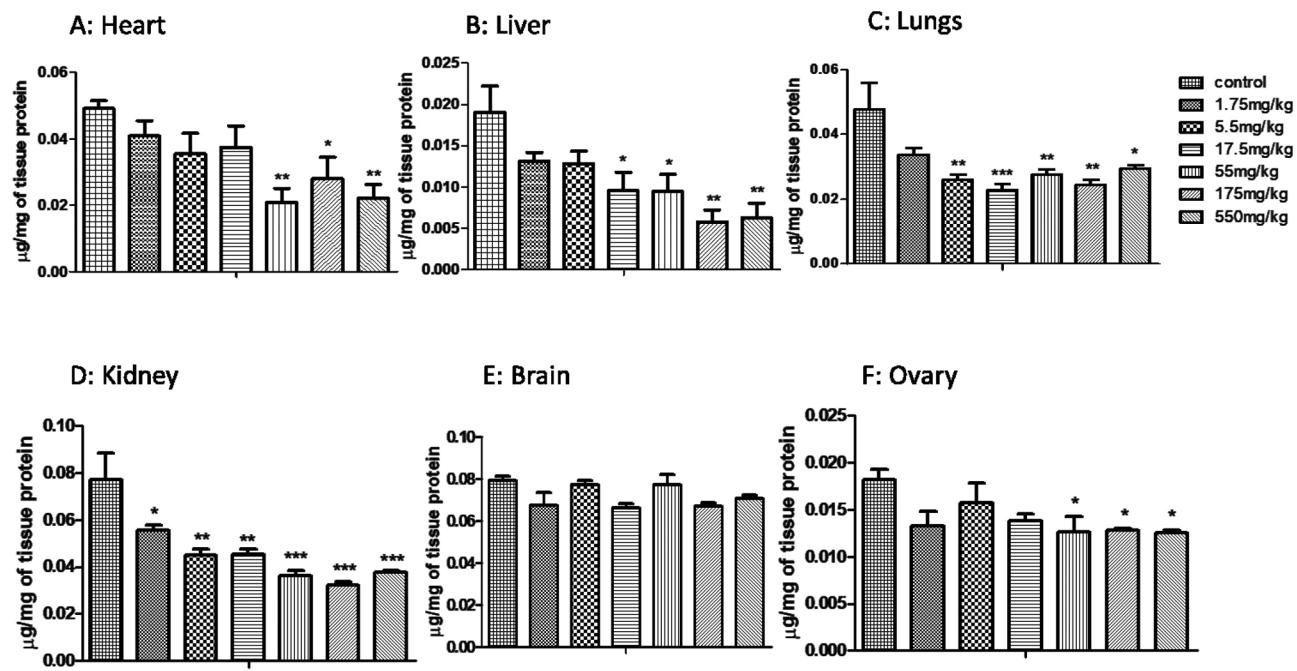
CAT levels were significantly decreased at both dose levels in both male and female treatment groups (Fig. 5). GSH levels were significantly decreased in both male and female treatment groups at both dose levels (Fig. 6), while MDA levels were increased in both treatment groups in all organs except brain (Fig. 7) SOD was also decreased at both doses in male and female rats groups as compared to normal control (Fig. 8). In the teratogenicity study, all the oxidative markers CAT, SOD and GSH were decreased while MDA levels were increased in the fetus when compared to the control (Fig. 9).

#### Effect of MoS<sub>2</sub> NF on biochemical markers in acute, sub-acute and developmental toxicity study

All the toxicity studies conducted in the present research showed a significant increase ( $p<0.001$ ) in ALT, AST levels in the liver (see Table 5 for acute, Table 7 for sub-acute and Table 8 for developmental toxicity study). RFTs showed a significant increase ( $p<0.001$ ) in Urea levels in acute toxicity study (Table 5) and female group of sub-acute toxicity study (Table 7) while the creatinine levels were significantly decreased ( $p<0.001$ ) in the same groups when compared to normal control. RFTs and lipid profile were comparable to normal control in developmental toxicity study (Table 9). In acute toxicity study, levels of cholesterol, LDL and Triglycerides were decreased while the HDL was increased ( $p<0.001$ ) (Table 6). Cholesterol level was increased ( $p<0.01$ ) in male treatment group while decreased ( $p<0.001$ ) in



**Fig. 1** Effect of  $\text{MoS}_2$ -NF (1.75 to 550 mg/kg) on Catalase levels in various tissues at the end of acute toxicity study. Data represented as Mean  $\pm$  SEM (n=5). \*\*\* $p$ <0.001 and \* $p$ <0.05 in comparison to control groups group

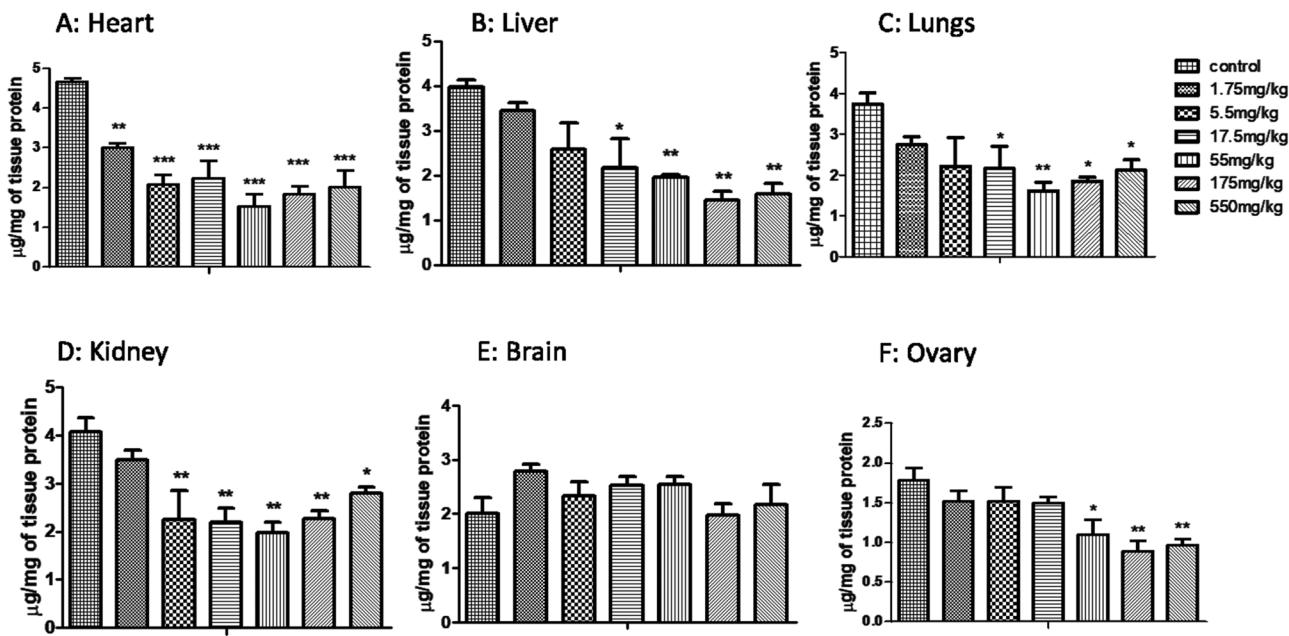


**Fig. 2** Effect of  $\text{MoS}_2$  NF treatment on GSH levels in acute toxicity studies of different organs at different logarithmic doses. Data represented as Mean  $\pm$  SEM (n=5). \*\*\* $p$ <0.001 and \* $p$ <0.05 in comparison to control groups group

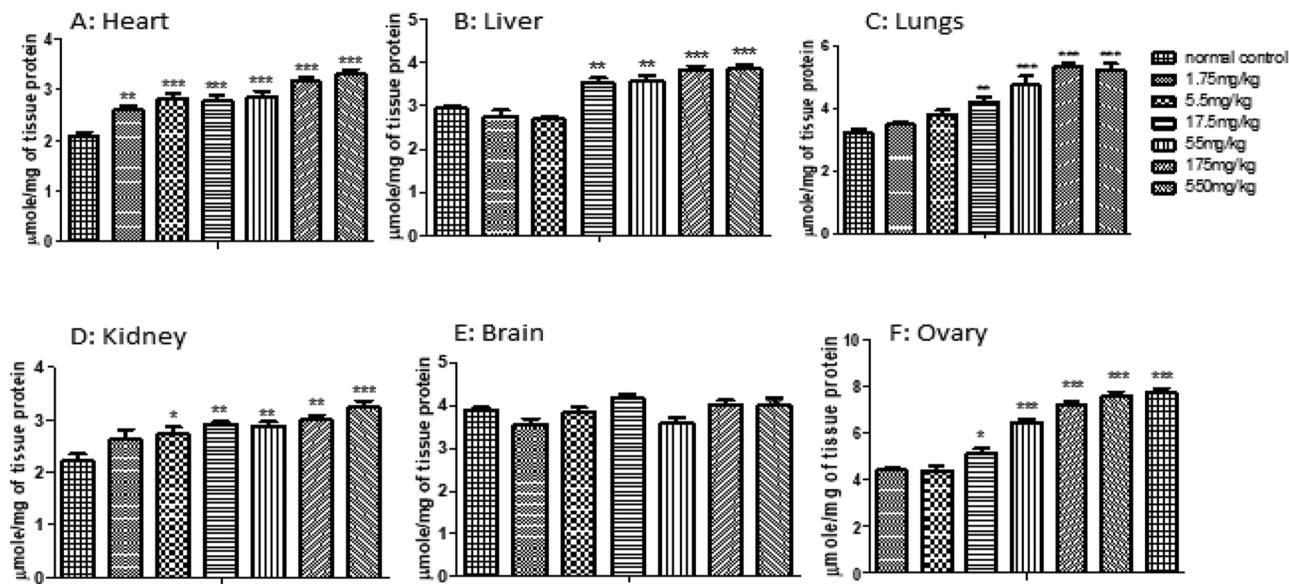
females. LDL was decreased ( $p$ <0.001) at low dose of 3 mg/kg while increased at high dose of 10 mg/kg  $\text{MoS}_2$  NF. Triglycerides were significantly increased at in both sex groups at both dose levels (Table 7).

#### Effect of sub-acute toxicity study on sperm count, motility, morphology and DNA integrity of sperms

Sperm concentration expressed an unexpected increase ( $p$ <0.001) at a dose of 10 mg/kg. Local motility of sperms appeared to rise ( $p$ <0.001) significantly at both doses but more at 3 mg/kg. Immotile sperms were notably increased ( $p$ <0.001) at 3 mg/kg dose of  $\text{MoS}_2$  NF while



**Fig. 3** Effect of  $\text{MoS}_2$  NF treatment on SOD levels in acute toxicity studies of different organs at different logarithmic doses. Data represented as Mean  $\pm$  SEM ( $n=5$ ). \*\*\* $p<0.001$ , \*\* $p<0.01$  and \* $p<0.05$  in comparison to control groups group



**Fig. 4** Effect of  $\text{MoS}_2$  NF treatment on MDA levels in acute toxicity studies of different organs at different logarithmic doses. Data represented as Mean  $\pm$  SEM ( $n=5$ ). \*\*\* $p<0.001$ , \*\* $p<0.01$  and \* $p<0.05$  in comparison to control groups group

10 mg/kg dose group of  $\text{MoS}_2$  NF was parallel to that of control group (Fig. 10). DNA integrity was not affected by the treatment (Fig. 11) while the sperms morphology was greatly affected (Fig. 11 and Table 8).

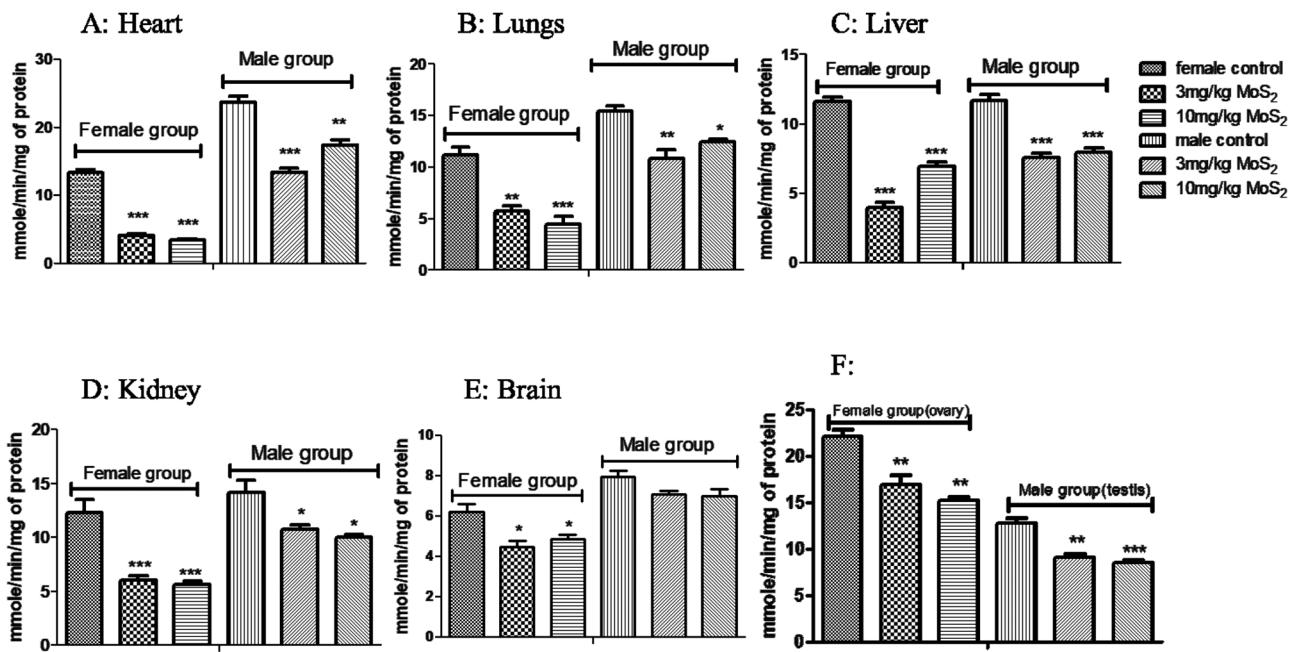
#### Measurement of teratogenic parameters after $\text{MoS}_2$ NF treatment

soft tissue examination showed normal growth of organs as compared to the control group (Fig. 12). The crown to rump length was found to be significantly increased

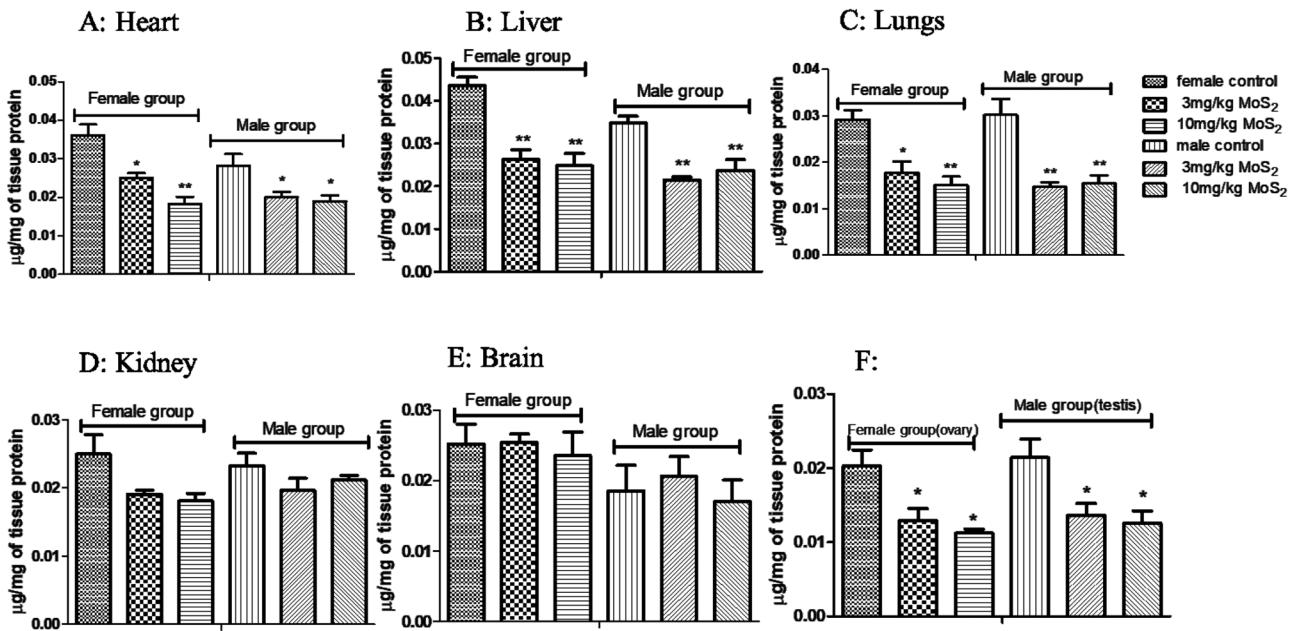
( $p<0.001$ ) when compared to normal control (Fig. 1 in supplementary data and Table 9). Weight of fetus was significantly increased while that of placental weight remained parallel to control (Table 9)

#### Effect of $\text{MoS}_2$ NF on histopathology of different organs in acute, sub-acute and developmental toxicity studies

Histopathology showed vacuolar degeneration in the brain in acute, developmental and sub acute study except in female at 3 mg/kg dose of subacute toxicity.



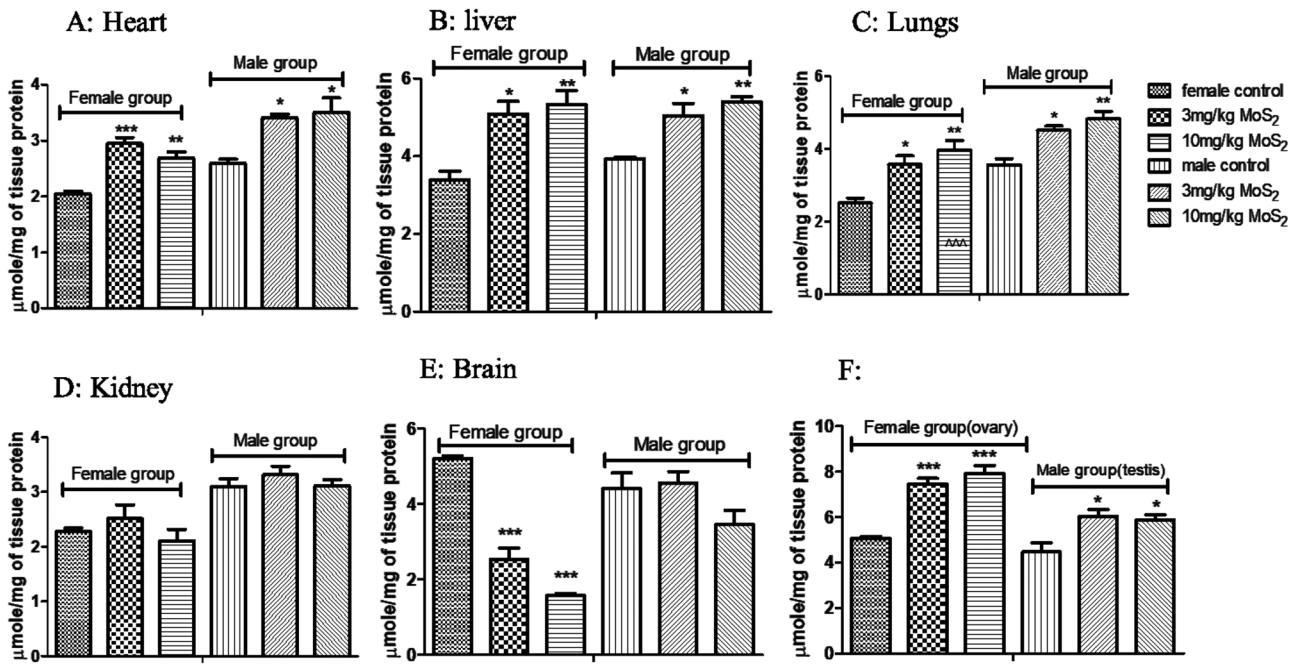
**Fig. 5** Effect of MoS<sub>2</sub>-NF (3 mg/kg and 10 mg/kg) on Catalase levels in various tissues at the end of sub-acute toxicity study. Data represented as Mean  $\pm$  SEM (n=5). \*\*\*p<0.001, \*\*p<0.01 and \*p<0.05 in comparison to control groups group



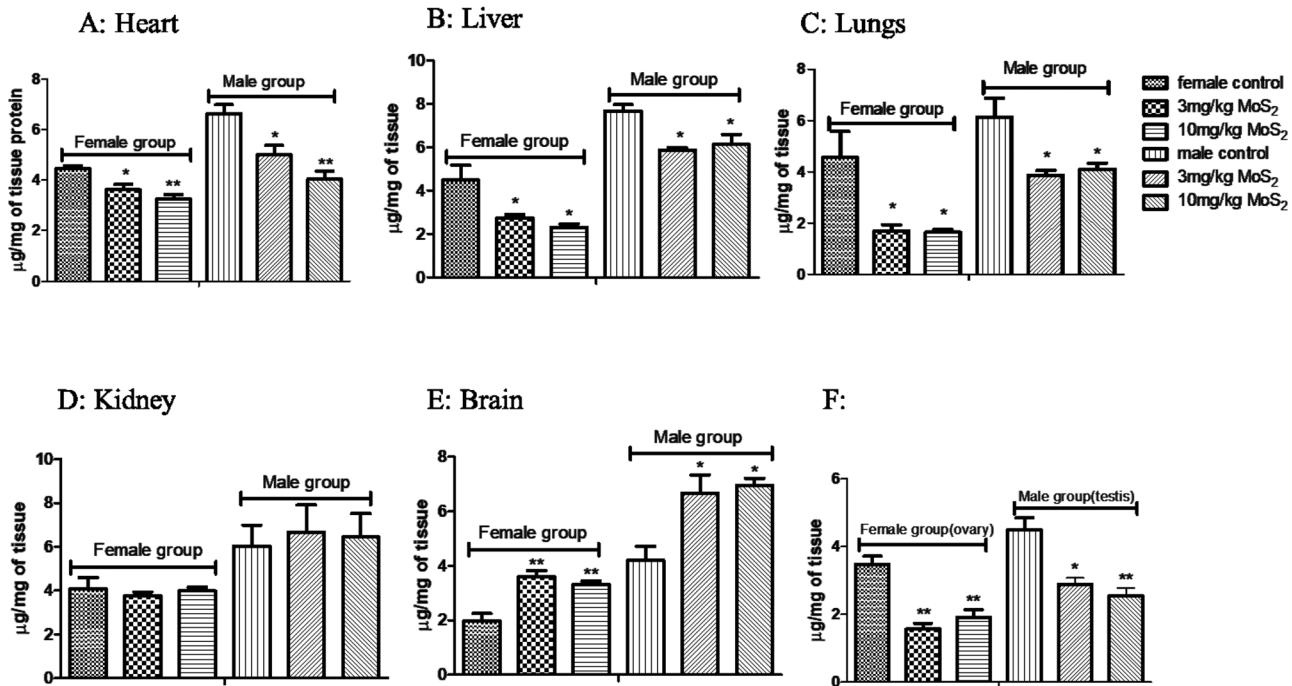
**Fig. 6** Effect of MoS<sub>2</sub>-NF (3 mg/kg and 10 mg/kg) on GSH levels in various tissues at the end of sub-acute toxicity study. Data represented as Mean  $\pm$  SEM (n=5). \*\*p<0.01 and \*p<0.05 in comparison to control groups group

Degeneration of cardiomyocyte in heart in all toxicity studies conducted. Degeneration in different regions of kidney and dilation of Bowman's capsule as well at all doses given in acute and sub-acute study (male group). Pyknosis and vascular congestion were also reported in the kidney at 10 mg/kg dose in the female treatment group in the subacute toxicity study. In lungs, Vascular

congestion was reported at the highest dose of 550 mg/kg and hemorrhage at 175 mg/kg dose. Histopathology of the liver showed hyaline degeneration in parenchyma cells in acute toxicity at a dose of 1.75 mg/kg and degenerative changes in the sinusoidal space at 5.5 mg/kg dose. Hepatocyte degeneration at dose 17.5 mg/kg, vascular congestion at 55 mg/kg dose level was seen in the



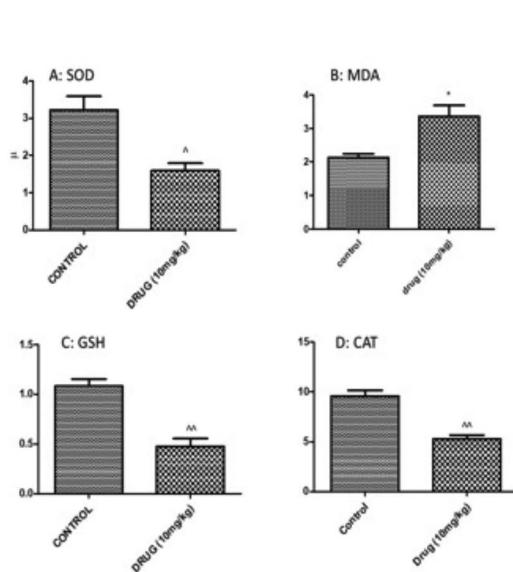
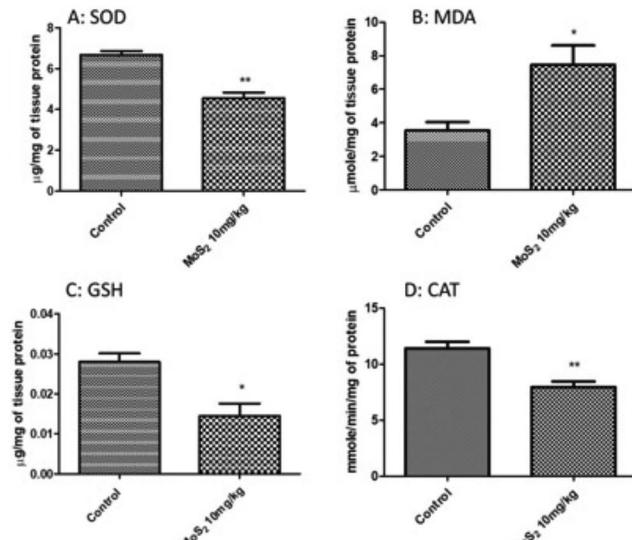
**Fig. 7** Effect of MoS<sub>2</sub>-NF (3 mg/kg and 10 mg/kg) on MDA levels in various tissues at the end of sub-acute toxicity study. Data represented as Mean  $\pm$  SEM (n=5). \*\*\*p<0.001, \*\*p<0.01 and \*p<0.05 in comparison to control groups group



**Fig. 8** Effect of MoS<sub>2</sub>-NF (3 mg/kg and 10 mg/kg) on SOD levels in various tissues at the end of sub-acute toxicity study. Data represented as Mean  $\pm$  SEM (n=5). \*\*\*p<0.001, \*\*p<0.01 and \*p<0.05 in comparison to control groups group

histopathological analysis of the liver. Swelling of hepatocytes was observed at 175 mg/kg in acute toxicity and at high dose (10 mg/kg) of sub-acute in female treatment group. Degenerative changes, pyknosis and necrosis were caused by the highest dose of 550 mg/kg in acute

toxicity. Degenerative changes were also reported in sub-acute toxicity study (Fig. 13). Swelling of hepatocytes was reported in developmental toxicity (Fig. 14) and sub-acute female 10 mg/kg group. Low doses of 1.75 and 5.5 mg/kg showed degeneration in ovarian parenchyma.

**A: Oxidative markers in pubs****B: Oxidative markers in Uterus of mothers****Fig. 9** Effect of MoS<sub>2</sub> NF 10 mg/kg on oxidative stress markers in pubs (A) and in uterus of mother (B) in developmental toxicity studies**Table 5** Effect of treatment on Liver and Renal functions in acute toxicity studies of MoS<sub>2</sub> NF at different logarithmic doses

Parameters	Unit	Normal control	MoS <sub>2</sub> NF	1.75 mg/kg	5.5 mg/kg	17.5 mg/kg	55 mg/kg	175 mg/kg	550 mg/kg
<b>LFTs</b>									
ALT	U/L	32.66 ± 1.20		87.0 ± 1.15***	41.33 ± 1.76*	50.33 ± 1.76***	77.33 ± 1.76***	73.66 ± 1.85***	108.0 ± 3.51***
AST	U/L	38.33 ± 2.02		135.66 ± 2.02***	106.66 ± 2.96***	107.66 ± 4.48***	261.0 ± 2.30***	215.0 ± 2.08***	208.66 ± 3.38***
Albumins	g/dl	3.53 ± 0.20		4.23 ± 0.176	3.50 ± 0.173	3.36 ± 0.20	3.76 ± 0.14	4.86 ± 0.14***	4.76 ± 0.145***
Globulins	g/dl	3.0 ± 0.11		3.16 ± 0.14	4.66 ± 0.14***	4.76 ± 0.14***	4.56 ± 0.145***	3.63 ± 0.176	3.76 ± 0.20*
<b>RFTs</b>									
Urea	mg/dL	37.66 ± 1.45		59.0 ± 1.15***	62.33 ± 1.76***	56.00 ± 2.64***	53.0 ± 2.08***	67.0 ± 1.52***	65.0 ± 2.08***
Creatinine	mg/dL	0.70 ± 0.012		0.61 ± 0.01	0.59 ± 0.02	0.48 ± 0.02	0.63 ± 0.01	0.55 ± 0.03	0.45 ± 0.03

Data are expressed as mean ± SEM, n=5, \*p<0.05, p<0.01, \*\*\*p<0.001 as compared with normal control. \*indicates increase levels while ^ indicate decreased levels

**Table 6** Effect of MoS<sub>2</sub> NF on lipid profile of different organs at logarithmic doses in acute toxicity studies

Parameters	Unit	Normal control	MoS <sub>2</sub> NF	group	1.75 mg/kg	5.5 mg/kg	17.5 mg/kg	55 mg/kg	175 mg/kg	550 mg/kg
Cholesterol	mg/dL	144.66 ± 1.45		group	90.33 ± 2.72^**	67.66 ± 3.38^**	72.00 ± 3.60^**	74.00 ± 3.05^**	77.33 ± 3.28^**	60.66 ± 2.96^**
Triglycerides	mg/dL	135.0 ± 1.73			108.66 ± 1.45	90.0 ± 2.64^**	103.66 ± 2.84^**	107.0 ± 4.58^**	98.0 ± 3.21^**	88.0 ± 3.0^**
LDL	mg/dL	75.33 ± 1.45			25.33 ± 1.76^**	34.0 ± 2.64^**	33.33 ± 1.76^**	44.33 ± 1.76^**	25.33 ± 2.33^**	24.66 ± 1.45^**
HDL	mg/dL	35.33 ± 2.02			50.0 ± 2.0***	48.66 ± 1.45***	50.0 ± 2.00***	48.0 ± 2.00**	46.5 ± 1.50**	52.0 ± 1.15***
VLDL	mg/dL	18.33 ± 0.88			20.33 ± 0.88	18.0 ± 1.15	18.66 ± 0.88	21.0 ± 1.15	20.0 ± 1.15	17.33 ± 0.88

Data are expressed as mean ± SEM, n=5, \*P<0.05, \*\*P<0.01, \*\*\*P<0.001 as compared with normal control. \*indicates increase levels while ^ indicate decreased levels

Cysts and numerous immature follicle structures were formed in the Ovaries in acute at doses of 17.5 and 55 mg/kg (Fig. 15) and sub-acute toxicity study (Fig. 13).

**Discussion**

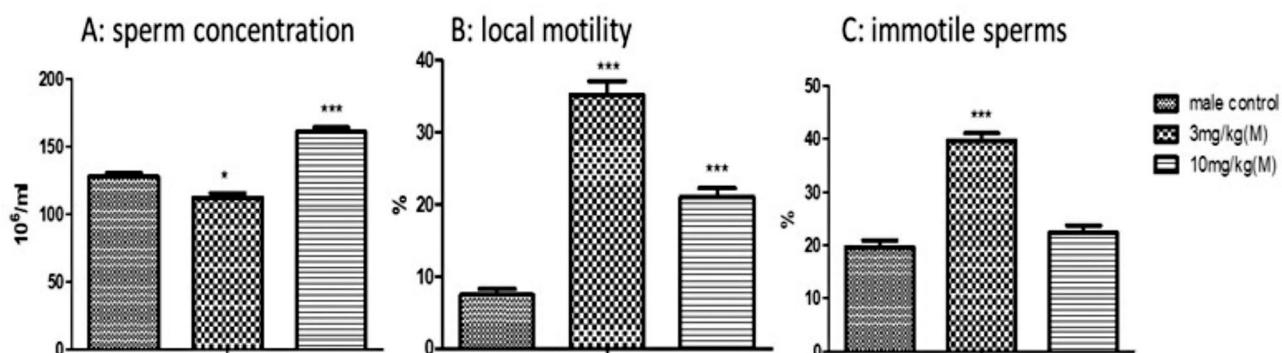
Prior to clinical trials, toxicity assessment is crucial in the development and screening of newly developed drugs. The primary goal of the toxicity study of any compound

is to make sure that the exposed population is secure from the toxic reactions of that compound [22]. International regulatory authorities for toxicity research (GSH, EPA, OECD, EEC etc.) can discover any possible hazards and risks of the substances on people by assessing the dose-dependent toxic outcomes that manifest in animal models [23].

**Table 7** Effect of MoS<sub>2</sub> on liver, renal function tests and lipid profile in sub-acute toxicity studies at 3 mg/kg and 10 mg/kg dose levels

Male		Control group	MoS <sub>2</sub> NF		Control group	Female	
LFTs	Units		3 mg/kg	10 mg/kg		MoS <sub>2</sub> NF	3 mg/kg
ALT	U/L	39.33±2.60	86.0±3.4***	56.0±3.2*	35.3±1.76	88.0±3.4***	58.33±3.48**
ALP	U/L	113.6±8.9	183.3±5.54**	216.3±10.4***	107.33±2.9	203.0±10.4**	414.0±16.0***
AST	U/L	38.3±2.3	147.3±7.8***	106.3±4.6***	31.33±1.76	135.33±4.91***	136.0±5.68***
<b>RFTs</b>							
Creatinine	mg/dL	0.53±0.06	0.60±0.05	0.68±0.04	0.76±0.03	0.61±0.04	0.53±0.03^
Urea	mg/dL	48.0±1.52	43.3±2.60	53.33±1.76	42.66±1.45	56.0±2.08**	51.0±2.30*
<b>Lipid Profile</b>							
Cholesterol	mg/dL	98.3±2.3	116.0±3.05**	90.6±2.9	105.3±1.7	93.6±3.5 ^	72.3±1.76^
Triglycerides	mg/dL	80.66±2.96	94.6±2.9*	97.0±3.05 *	88.6±2.02	114.3±3.2 **	104.0±3.60*
HDL	mg/dL	48.3±1.45	50.0±2.08	49.0±1.73	45.6±1.4	51.3±1.76	53.3±2.3*
LDL	mg/dL	48.6±2.02	46.0±2.8	49.6±2.02	63.6±2.02	28.6±2.02^	81.0±2.6**
VLDL	mg/dL	19.0±1.15	17.3±0.8	20.0±1.15	24.0±1.15	20.6±0.8	23.6±1.45

Data are expressed as mean±SEM, n=5, \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 as compared with normal control. ^indicates increase levels while ^ indicate decreased levels

**Fig. 10** Effect of treatment on sperm Motility and Concentration in Sub-acute toxicity. \*\*\*p<0.001 and \*p<0.05 in comparison to control group

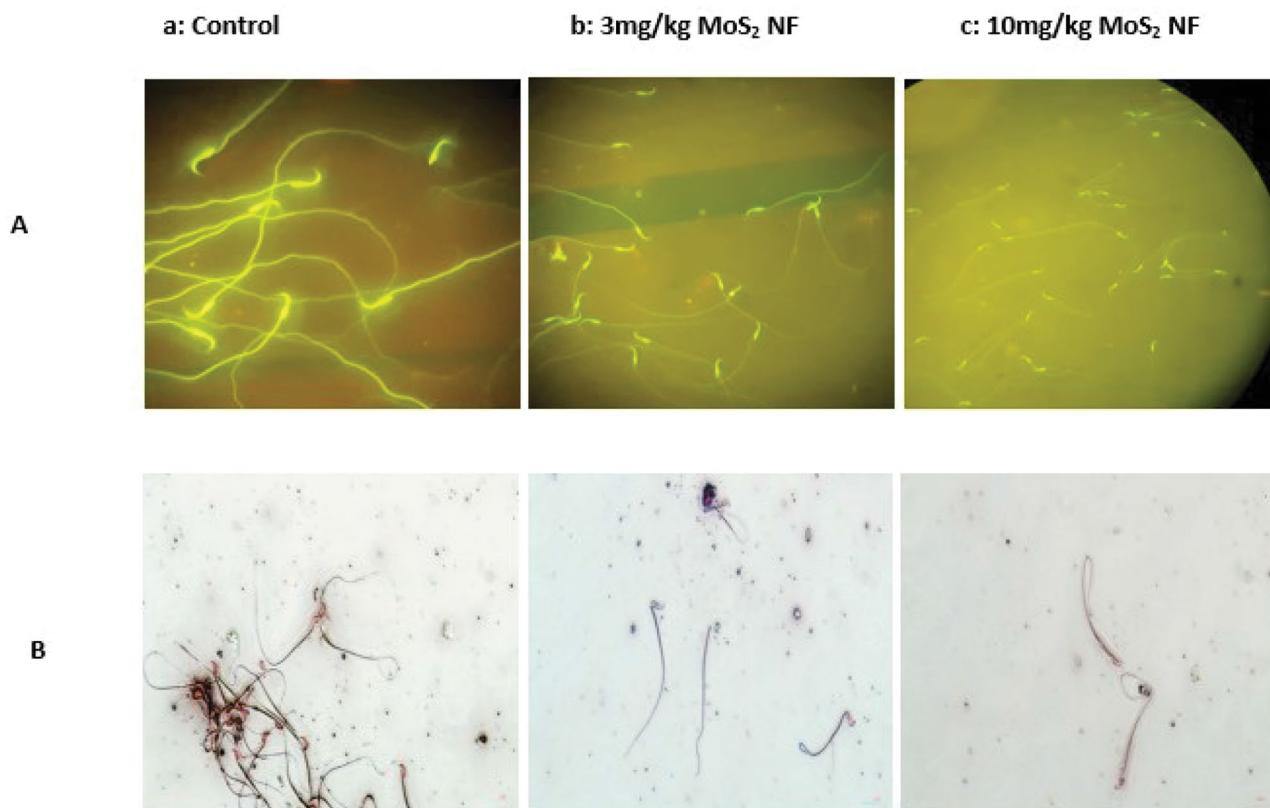
In acute oral toxicity studies, a single drug dosage is used to determine lethal dose (LD<sub>50</sub>). In this study, main test was conducted using logarithmic doses of 1.75 mg/kg to 550 mg/kg of MoS<sub>2</sub> Nano flowers following the OECD guidelines. This was done because mostly toxicities studies are conducted at high doses, but humans have a less probability of getting large doses so this study was aimed at checking the toxicity at low doses of metal NF [24]. Both the acute and sub-acute toxicity investigation showed no evidence of mortality at any dose.

Body weights were significantly decreased during the 14-day study duration of acute toxicity studies and were comparable to control in sub-acute toxicity studies, while in developmental toxicity studies fetal weights were increased and maternal weights were comparable to normal control. Organ weights were increased in acute toxicity studies and sub-acute male treatment group. This increase and decrease in body weights might be an

indication of adverse drug reaction or it might be due to improved or healthy diet [11, 25].

Changes in liver and renal function parameters might be associated with their toxicity, as they are the major sites for metabolism and elimination of substances. Increase in ALT and AST levels may be due to liver injury that makes the membrane more permeable and causes the release of aminotransferases in the blood [26–28]. In the current research, ALT, AST, globulins and albumin levels were increased in all the three types acute, sub-acute and developmental toxicity studies. Previous studies revealed that increase in ALT, AST levels are indicative of hepatocellular damage, so MoS<sub>2</sub> NF might be hepatotoxic [25, 29, 30].

Renal function remains comparable to normal control in all study groups except in the female treatment groups of sub-acute toxicity studies where blood urea level was increased at both dose levels which in accordance to the

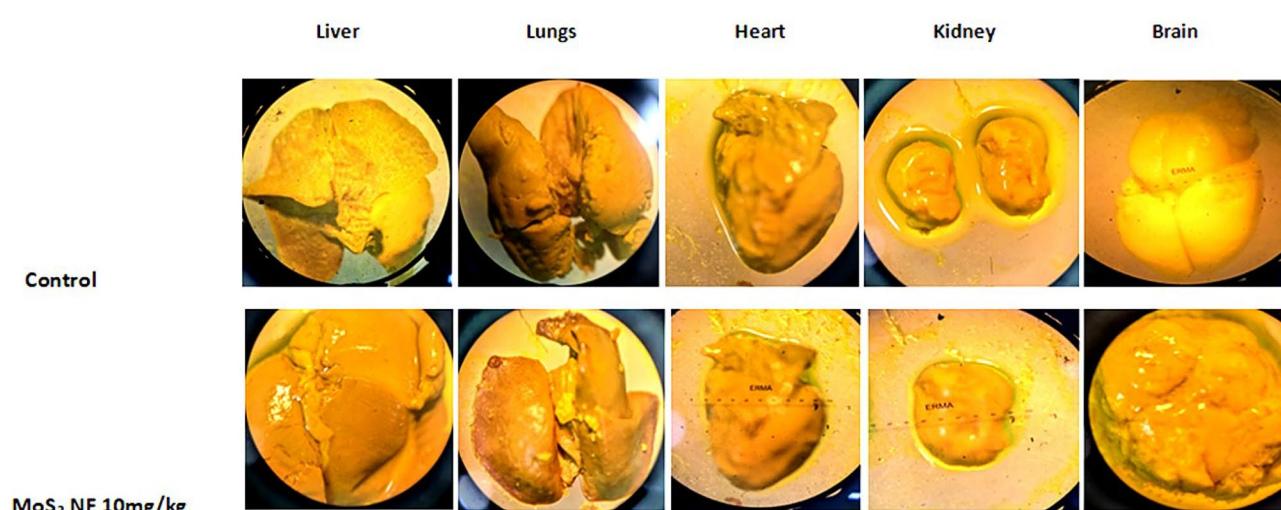


**Fig. 11** Effect of treatment on sperm's DNA integrity (A) and morphology (B) in Sub-Acute toxicity study

**Table 8** Effect of MoS<sub>2</sub> NF on sperm morphology in sub-acute toxicity studies

Parameters	Sperm defects					
	Headless (%)	Hookless (%)	Curved tail (%)	Bend (%)	Coil tail (%)	Normal (%)
CMC	9	1.0	8	6.2	5.3	70.5
MoS <sub>2</sub> NF (3 mg/kg)	13*	1.3	12.2	8.3*	8.0	57.2 <sup>▲▲</sup>
MoS <sub>2</sub> NF (10 mg/kg)	20***	1.5*	14.5**	10.56***	12.0**	41.44 <sup>▲▲</sup>

Data are expressed as mean  $\pm$  SEM, n=5, \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 as compared to control. \*indicates increase levels while ^ indicate decreased levels



**Fig. 12** Effect of MoS<sub>2</sub> NF on pups' organs in developmental toxicity study

**Table 9** Effect of MoS<sub>2</sub> NF on liver, renal functions, lipid profile, crown to rump length, placental and fetal weight of animals in developmental toxicity studies

LFTs	Units	Control group (CMC)	MoS <sub>2</sub> NF (10 mg/kg)
Bilirubin	mg/dL	0.750±0.076	0.533±0.145
ALT	U/L	41.67±2.02	106.7±7.83**
AST	U/L	35.33±4.33	189.3±8.25***
ALP	U/L	142.3±3.38	115.0±11.55
Globulins	g/dL	3.55±0.24	2.93±0.24
<b>RFTs</b>			
Urea	mg/dL	43.67±2.96	45.33±5.23
Creatinine	mg/dL	0.873±0.063	0.626±0.093
<b>Lipid Profile</b>			
Cholesterol	mg/dL	111.0±3.0	118.0±3.0
Triglycerides	mg/dL	93.0±3.00	106.0±4.00
HDL	mg/dL	47.00±2.00	48.50±2.50
LDL	mg/dL	61.00±2.00	53.50±2.50
VLDL	mg/dL	22.00±2.00	24.50±1.50
<b>Crown rump length</b>	cm	4.760±0.242	6.060±0.06782***
<b>Placental weight variation</b>	g	0.7240±0.1256	0.6480±0.04510
<b>Fetal weight variation</b>	g	2.296±0.136	5.286±0.2685***

Data are expressed as mean±SEM, n=5, \*\*P<0.01, \*\*\*P<0.001 as compared to normal control. \*indicates increase levels while ^indicate decreased levels

previous studies are due to renal damage because most of the drugs are excreted by kidney [31]

Lipid profile of the acute oral toxicity study exhibited a remarkable decrease that indicates MoS<sub>2</sub> might be hypolipidemic [31]. Lipid profile assessment of sub-acute toxicity studies showed increased levels of triglycerides in both sex groups at all dose levels, which in accordance with previous studies might be a cause of cardiovascular disease [32]. Cholesterol level was increased in 3 mg/kg MoS<sub>2</sub> NF group while decrease levels were observed in all female treatment groups. Variation in cholesterol levels are related to hypo and hyperlipidemia. In the male treatment group, LDL was decreased at low dose while increase at high dose of 10 mg/kg MoS<sub>2</sub> NF with a notable increase in HDL level at the same dosage [31]. In developmental toxicity studies, lipid profile remains intact.

Hematological assay of the acute toxicity study showed increased levels of hemoglobin, RBCs WBCs and neutrophils at high logarithmic doses while the lymphocyte count was decreased at the high dose level. This rise might be an immune-protective effect in response to the toxic outcomes of the medicine [33]. Hematological

analysis of sub-acute toxicity assessment showed an increase in RBC in female 10 mg/kg MoS<sub>2</sub> NF treatment group which might be due to low oxygen level in the blood circulation which in turn stimulate erythropoietin secretion in the kidney [32]. Neutrophils were decreased in the sub-acute and developmental toxicity investigation while lymphocytes were increased in sub-acute female 3 mg/kg MoS<sub>2</sub> NF treatment group and developmental toxicity and decreased in all male treatment groups. Platelet count was increased in male treatment group of sub-acute toxicity studies and developmental toxicity studies. According to previous toxicities studies, this increase or decrease might be due to immune response [11].

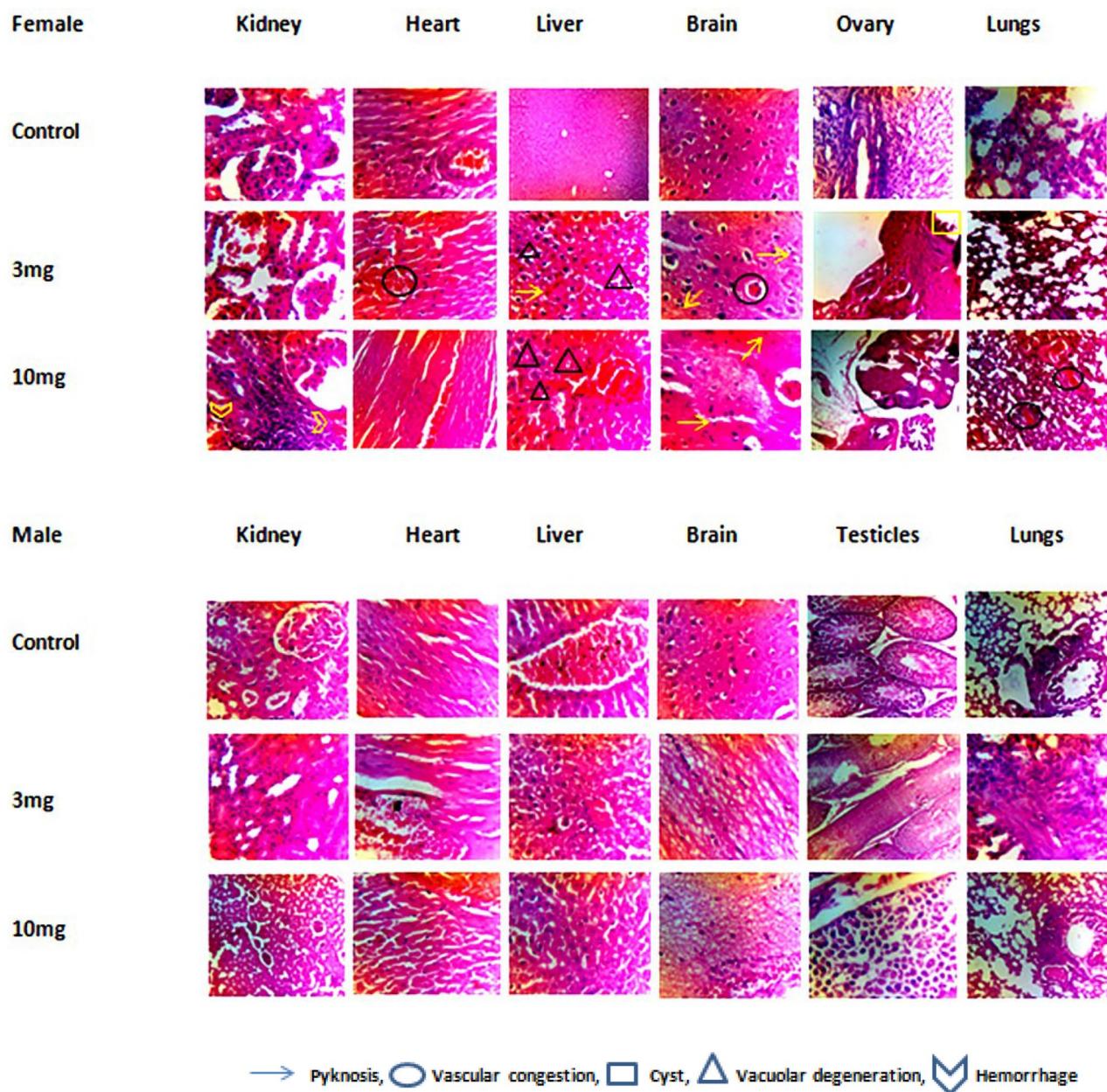
During toxicity, ROS are responsible for creating oxidative stress pathways. They are usually responsible for cell signaling and homeostasis [34]. Disease state, toxins and radiation exposure are the factors that lead to an abnormal elevation in ROS [16]. Prolonged exposure or production of reactive oxygen species in organisms may result in altered oxidative balance, which can harm lipids, proteins and nucleic acid [35]. It can cause apoptosis, necrosis and irreparable cell damage to the affected cell. A number of defense mechanism pathways are then activated to protect against these reactive oxygen species [11]. Antioxidant enzymes, including SOD, CAT, and GSH, are involved in this defense mechanism.

In the acute toxicity studies CAT, SOD and GSH showed decrease levels in all the selected organs except brain at almost all doses that might indicate antioxidant property of MoS<sub>2</sub> NF.

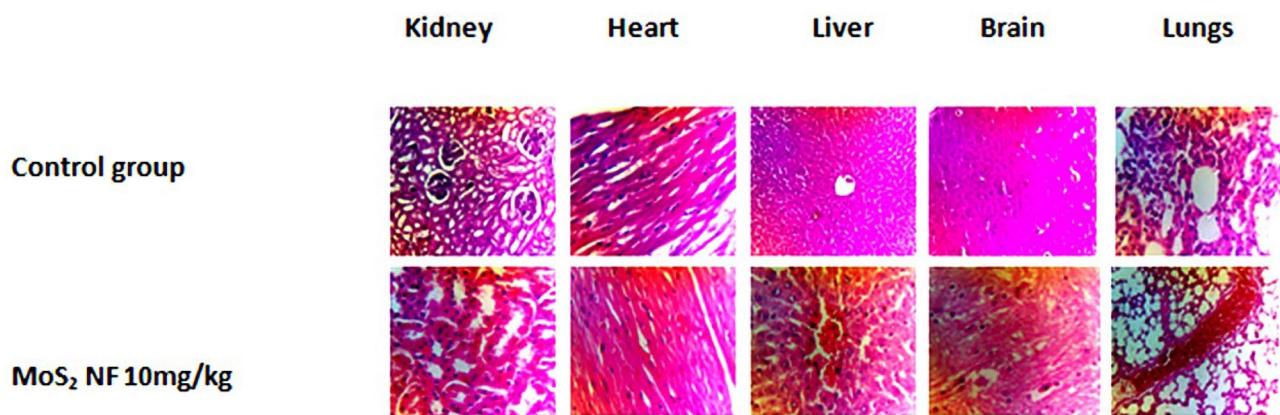
MDA levels showed increased activity except Brain. This increase might indicate potential oxidative stress and harm to tissues [11]

In sub-acute toxicity research SOD, CAT, GSH and MDA were measured in different tissues to check for any change or cell damage caused by MoS<sub>2</sub> NF. CAT, SOD and GSH levels were significantly decreased at all dose levels in both male and female treatment groups while MDA levels were increased. In teratogenicity study all the oxidative markers were decreased in fetus except MDA which was increased which might show the toxic potential of the drug.

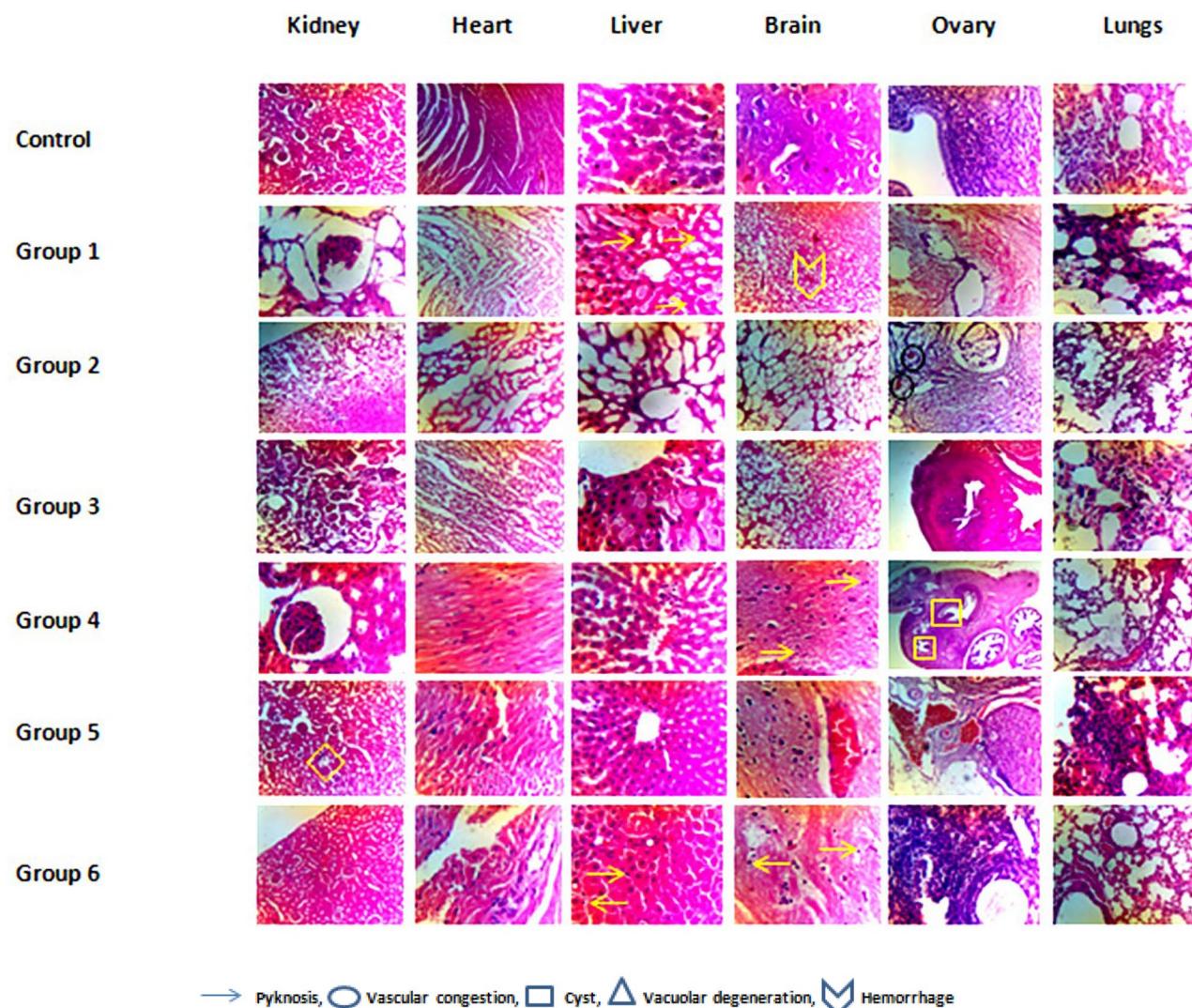
DNA integrity was analyzed in male treatment groups of sub-acute toxicity as it is the major indicator of fertility and is strongly linked to mobility of sperm; a parameter for sperm functioning. In the ongoing research, DNA integrity of the sperms was normal, indicating that Blood-testis barrier is not affected by the drug [13].



**Fig. 13** Histopathological analysis of male and female rats' organs in sub-acute toxicity



**Fig. 14** Represents histopathological changes of the maternal organs in developmental toxicity



**Fig. 15** Histopathology of different organs in acute toxicity study

Sperm count was increased at a high dose of 10 mg/kg and was decreased at a low dose. Sperm motility was increased at both dose levels. Immotile sperms increased at low dose while comparable to control at high dose. Histopathology of different organs in all the three toxicity studies performed was altered by drug treatment.

## Conclusion

This study provide a comprehensive toxicological assessment of the (MoS<sub>2</sub> NFs) through acute, sub-acute, and developmental toxicity studies in Wistar rats, following OECD guidelines. The finding demonstrates that MoS<sub>2</sub> NF exhibit mild-to-moderate level toxicity, characterized by hepatic and renal dysfunction, oxidative stress induction, hematological alterations, and histopathological changes in vital organs. The observed toxicities are likely mediated by altered liver enzymes, hematological disturbances, oxidative stress and histopathological changes, including cellular degeneration and inflammatory responses. Although MoS<sub>2</sub> NFs did not cause mortality at the tested doses, the histopathological changes, alterations in biochemical markers, and oxidative stress induction suggest potential risks associated with prolonged exposure.

However, further studies are essential to ensure the safe and effective utilization of MoS<sub>2</sub> NFs in therapeutic applications.

## Abbreviations

ALT	Alkaline transaminase
ALP	Alkaline phosphatase
AST	Aspartate Transaminase
ANOVA	Analysis of Variance
CAT	Catalase
CBC	Complete blood count
CMC	Carboxymethyl cellulose
DNA	Deoxyribonucleic Acid
EDTA	Ethylene diamine tetra acetic acid
GSH	Glutathione
HDL	High density lipoprotein
LD	Lethal dose
LDL	Low density lipoprotein
VLDL	Very low density lipoprotein
LFTs	Liver function test
MoS <sub>2</sub> NF	Molybdenum disulfide Nanoflowers
MDA	Malonaldehyde
OECD	Organization for Economic Co-operation and Development
RFTs	Renal function test
RBCs	Red blood cells
ROS	Reactive oxygen species
SOD	Superoxide dismutase
SEM	Standard error of Mean
WBCs	White blood cells

## Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s40360-025-00881-8>.

Supplementary Material 1

## Acknowledgements

Not Applicable

## Author contributions

FH carried out experimental work and prepared the first draft of the manuscript. AK designed and supervised the project, prepared and revised the manuscript, and conducted data analysis. FA Designed and supervised the research project; helped in experimental work and data analysis. HU Synthesized, characterized and provided the MoS<sub>2</sub>NF. MFA and MIK helped in the revision of the manuscript, data analysis, and critical revision of the drafted manuscript. All authors approved the final version of the manuscript.

## Funding

No funding is provided for this study.

## Data availability

Data is provided within the manuscript or supplementary information files.

## Declaration

### Ethical approval

All the procedures and techniques used in this study were approved by the Ethical committee (REC/RIPS/LHR/2023/068) of Riphah International University, Raiwind Campus, Lahore, Pakistan.

### Consent to participate and consent for publication

Not applicable

### Competing interests

The authors declare no competing interests.

Received: 28 October 2024 / Accepted: 25 February 2025

Published online: 25 August 2025

## References

1. Hullu J, Sahu S, Hayes A. Nanotechnology: history and future. *Hum Exp Toxicol.* 2015;34(12):1318–21.
2. Nikzamir M, Akbarzadeh A, Panahi Y. Technology. An overview on nanoparticles used in biomedicine and their cytotoxicity. 2021;61:102316.
3. Odularu AT, Ajibade PA, Mbese JZ. Impact of molybdenum compounds as anticancer agents. *Bioinorg Chem Appl.* 2019;2019.
4. Barceloux DG, Barceloux D. Molybdenum. *J Toxicol Clin Toxicol.* 1999;37(2):231–37.
5. Benavente E, Santa Ana M, Mendizábal F, González G. Intercalation chemistry of molybdenum disulfide. *Coord Chem Rev.* 2002;224(1–2):87–109.
6. Vilian A, Dinesh B, Kang S-M, Krishnan UM, Huh YS, Han Y-K. Recent advances in molybdenum disulfide-based electrode materials for electroanalytical applications. *Mikrochim Acta.* 2019;186(3):1–29.
7. Li J, Guiney LM, Downing JR, Wang X, Chang CH, Jiang J, et al. Dissolution of 2D Molybdenum disulfide generates differential toxicity among liver cell types compared to Non-Toxic 2D Boron Nitride effects. *Small* 2021;17:2101084.
8. Roheel A, Khan A, Anwar F, Ullah H, Rehman AU, Ullah N, et al. Evaluation of anti-tumor activity of molybdenum disulfide nanoflowers per se and in combination with berberine against mammary gland cancer in rats. *J Nanopart Res* 2024;26(10):1–24. <https://doi.org/10.1007/s11051-024-06153-6> <https://link.springer.com/article/10.1007/s11051-024-06153-6>
9. Guideline OO. 425: acute oral toxicity—up-and-down procedure. OECD Guidelines for the Test Chem. 2001;2:12–16.
10. Anwar F, Saleem U, Ahmad B, Ismail T, Mirza MU, Ahmad S. Acute oral, subacute, and developmental toxicity profiling of naphthalene 2-yl, 2-chloro, 5-nitrobenzoate: assessment based on stress response, toxicity, and adverse outcome pathways. *Front Pharmacol.* 2022;12:3967.
11. Anwar F, Saleem U, Rehman A-U, Ahmad B, Freyen M, Mirza MU, et al. Toxicity evaluation of the naphthalen-2-yl 3, 5-dinitrobenzoate: a drug candidate for alzheimer disease. *Front Pharmacol.* 2021;12:607026.
12. Kunimatsu T, Yamada T, Miyata K, Yabushita S, Seki T, Okuno Y, et al. Evaluation for reliability and feasibility of the draft protocol for the enhanced rat 28-day

subacute study (OECD Guideline 407) using androgen antagonist flutamide. *Toxicology* 2004;200:77–89.

13. Saleem U, Zubair S, Riaz A, Anwar F, Ahmad B. Effect of venlafaxine, pramipexole, and valsartan on spermatogenesis in male rats. *ACS Omega*. 2020;5(32):20481–90.
14. Anwar F, Saleem U, Rehman AU, Ahmad B, Ismail T, Mirza MU, et al. Toxicological screening of 4-Phenyl-3, 4-dihydrobenzo [h] Quinolin-2 (1 H)-one: a new potential candidate for Alzheimer's treatment. *ACS Omega* 2021;6:10897–909.
15. El-Boshy ME, Risha EF, Abdelhamid FM, Mubarak MS, Hadda TB. Protective effects of selenium against cadmium induced hematological disturbances, immunosuppressive, oxidative stress and hepatorenal damage in rats. *J Trace Elem Med Biol.* 2015;29:104–10.
16. Anwar F, Saleem U, Ahmad B, Ismail T, Mirza MU, Ahmad S. Acute oral, subacute, and developmental toxicity profiling of naphthalene 2-yl, 2-chloro, 5-nitrobenzoate: assessment based on stress response, toxicity, and adverse outcome pathways. *Front Pharmacol.* 2022;12:810704.
17. Saleem U, Ahmad B, Ahmad M, Hussain K, Bukhari NI. Investigation of in vivo antioxidant activity of *Euphorbia helioscopia* latex and leaves methanol extract: a target against oxidative stress induced toxicity. *Asian Pac J Trop Med.* 2014;7:S369–S75.
18. Parambi DGT, Saleem U, Shah MA, Anwar F, Ahmad B, Manzar A, et al. Exploring the therapeutic potentials of highly selective oxygenated chalcone based MAO-B inhibitors in a haloperidol-induced murine model of Parkinson's disease. *Neurochem Res.* 2020;45:2786–99.
19. Mir NT, Saleem U, Anwar F, Ahmad B, Ullah I, Hira S, et al. *Lawsonia inermis* markedly improves cognitive functions in animal models and modulate oxidative stress markers in the brain. *Medicina* 2019;55:192.
20. Bhangale JO, Acharya SR. Anti-Parkinson activity of petroleum ether extract of *Ficus religiosa* (L.) leaves. *Adv Pharmacol Pharma Sci.* 2016;2016.
21. Saleem S, Anwar F, Khan A, Saleem U, Akhtar MF, Shahzadi I, et al. Toxicity profiling of *Burgmansia aurea* Lagerh. Leaves using acute and sub-acute toxicity studies in rats. *J Ethnopharmacol.* 2023;311:116447.
22. Morton DM. Importance of species selection in drug toxicity testing. *Toxicol Lett.* 1998;102:545–50.
23. Lewis RW, Billington R, Debryune E, Gamer A, Lang B, Carpanini F. Recognition of adverse and nonadverse effects in toxicity studies. *Toxicol Pathol.* 2002;30(1):66–74.
24. Kaur J, Khatri M, Puri S. Toxicological evaluation of metal oxide nanoparticles and mixed exposures at low doses using zebra fish and THP1 cell line. *Environ Toxicol.* 2019;34(4):375–87.
25. Mounnissamy VM, Kavimani S, Sankari G, Quine SD, Subramani K. Evaluation of acute and sub-acute toxicity of ethanol extracts of *Cansjera rheedii* J. Gmelin (Opiaceae). *J Brew Distilling.* 2010;1(1):011–4.
26. Ain QU, Saleem U, Ahmad B, Khalid I. Pharmacological screening of silibinin for antischizophrenic activity along with its acute toxicity evaluation in experimental animals. *Front Pharmacol.* 2023;14:1111915.
27. Ogunkana OO, Ogunkana OE, Adeneye AA, Udo-Chijioke O, Dare-Olpede T, Olagunju JA, et al. Evaluation of the toxicological profile of the leaves and young twigs of *Caesalpinia bonduc* (Linn) roxb. *Afr J Traditional, Complementary Altern Med* 2013;10:504–12.
28. Kasarala G, Tillmann HL. Standard liver tests. *Clin Liver Dis.* 2016;8(1):13–18.
29. Olaniyan JM, Muhammad HL, Makun HA, Busari MB, Abdullah AS. Acute and sub-acute toxicity studies of aqueous and methanol extracts of *Nelsonia campestris* in rats. *J Acute Dis.* 2016;5(1):62–70.
30. Khan SM, Kour G. Subacute oral toxicity of chlorpyriphos and protective effect of green tea extract. *Pesticide Biochem Physiol.* 2007;89(2):118–23.
31. Hosseinzadeh H, Shakib SS, Sameni AK, Taghiabadi E. Acute and subacute toxicity of safranal, a constituent of saffron, in mice and rats. *Iran J Pharm Res.* 2013;12(1):93.
32. Magaye RR, Yue X, Zou B, Shi H, Yu H, Liu K, et al. Acute toxicity of nickel nanoparticles in rats after intravenous injection. *Int J Nanomed.* 2014;1393–402.
33. Mythilypriya R, Shanthi P, Sachdanandam P. Oral acute and subacute toxicity studies with *Kalpaamruttha*, a modified indigenous preparation, on rats. *J Health Sci.* 2007;53(4):351–58.
34. Schieber M, Chandel NS. ROS function in redox signaling and oxidative stress. *Curr Biol.* 2014;24(10):R453–R62.
35. Lushchak VI. Free radicals, reactive oxygen species, oxidative stress and its classification. *Chem Biol Interact.* 2014;224:164–75.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.